

MINIREVIEW

# Rimonabant: Just an Antiobesity Drug? Current Evidence on Its Pleiotropic Effects

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## ABSTRACT

The advent of the highly selective cannabinoid receptor (CB1) antagonist, rimonabant (SR141716; Acomplia) can revolutionize the ability of the clinicians to manage obesity. Large-scale clinical trials have demonstrated that rimonabant therapy can reduce obesity. Although, the precise mechanisms of action of rimonabant have to be further dissected, it is emerging, from

both preclinical and clinical research, that not only is rimonabant an antiobesity drug, but also its pleiotropic functions affect a broad range of diseases, from obesity-related comorbidities to drug dependence and cancer. Here we review recent data from the literature and discuss the full pharmacological potential of this drug.

Studies on the effect of marijuana psychoactive principle  $\Delta^9$ -tetrahydrocannabinol (THC) have evolved into the discovery and description of the endocannabinoid system. So far, this system is composed of two receptors (the widely expressed CB1 and the more restricted CB2), five endogenous lipid-like ligands [including the well known endocannabinoids anandamide (AEA) and 2-arachidonoyl glycerol], and the enzymes involved in their biosynthesis and degradation (for review, see Mechoulam et al., 1998; De Petrocellis et al., 2004; Di Marzo et al., 2004). Starting from the discovery of the endocannabinoid system, a number of studies have pointed out that altered endocannabinoid signaling and CB1 receptor expression are involved in several pathophysiological situations, ranging from neurological and psychiatric dis-

eases to eating, cardiovascular, and reproductive disorders. More recently, it has been described that CB1 receptor stimulation by the endocannabinoid AEA can negatively modulate cancer cell proliferation in vitro (Bifulco et al., 2001, 2004) as well as tumor growth and metastatic spreading in vivo (Portella et al., 2003; Bifulco et al., 2006, 2007). CB1 or CB2 antagonistic or inverse agonistic compounds have been used to investigate the endocannabinoid network and its integration with other signaling transduction pathways (for review, see Lange and Kruse, 2005). The first highly selective CB1 receptor antagonist, rimonabant (SR141716; Acomplia) was discovered by Sanofi-Aventis (Bridgewater, NJ) (Rinaldi-Carmona et al., 1994). It showed a number of biological effects in vitro and in vivo in several pathological situations. An update on the pleiotropic effects of rimonabant does not exist, whereas the knowledge concerning endocannabinoid system has been expanded considerably; therefore, we critically analyze the current literature on the pharmacological potential of rimonabant. We aim to describe both

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**ABBREVIATIONS:** THC,  $\Delta^9$ -tetrahydrocannabinol; AEA, anandamide (*N*-arachidonylethanolamine); SR141716, *N*-(piperidino-1-yl)-5-(4-chlorophenyl)-1-(2,4-dichlorophenyl)-4-methyl-pyrazole-3-carboxamide; CP 55,940, (1*R*,3*R*,4*R*)-3-[2-hydroxy-4-(1,1-dimethylheptyl)phenyl]-4-(3-hydroxypropyl)cyclohexan-1-ol; WIN 55,212-2, (*R*)-(+)-[2,3-dihydro-5-methyl-3-(4-morpholinylmethyl) pyrrolo-[1,2,3-*d,e*]-1,4-benzoxazin-6-yl]-1-naphthalenyl-methanone; MAPK, mitogen-activated protein kinase; HDL, high-density lipoprotein; RIO, rimonabant in obesity; DA, dopamine; msP, Marchigian Sardinian alcohol preferring rats; SR144528, *N*-[(1*S*)-endo-1,3,3-trimethyl bicyclo heptan-2-yl]-5-(4-chloro-3-methylphenyl)-1-(4-methylbenzyl)-pyrazole-3-carboxamide; LPS, lipopolysaccharide; TNF, tumor necrosis factor; MK-0364, *N*-[(1*S*,2*S*)-3-(4-chlorophenyl)-2-(3-cyanophenyl)-1-methylpropyl]-2-methyl-2-[[5(trifluoromethyl)pyridin-2-yl]oxy]propanamide; SR147778, [5-(4-bromophenyl)-1-(2,4-dichlorophenyl)-4-ethyl-*N*-(1-piperidinyl)-1*H*-pyrazole-3-carboxamide].

the clinical efficacy and the biological activity of rimonabant examining as much as possible the molecular aspects at the basis of rimonabant-induced effects. First, we discuss the efficacy and safety of rimonabant in reducing body weight and cardiometabolic risk factors. Second, we review the increasing literature on the other potential therapeutic properties of the CB1 receptor blocker rimonabant on behavior and in disorders related to the central nervous system. Third, we examine recent results about the antiproliferative effects of rimonabant. Finally, we discuss current data on rimonabant action as a modulator of reproductive system functions.

### In Vitro and in Vivo Pharmacology of Rimonabant

The discovery of endocannabinoid system prompted the development of CB1- and CB2-selective antagonists, the first of which was the CB1-selective rimonabant (SR141716) (Fig. 1). Binding studies have demonstrated that rimonabant is a potent ( $pK_i = 8.4$ ) and selective ligand for CB1 receptors, showing a high affinity ( $K_i = 5.6$  nM) for the CB1, and low affinity ( $K_i > 1000$  nM) for the CB2 receptor (Rinaldi-Carmona et al., 1994). Moreover, it displays a weak affinity to Galanin<sub>2</sub>, MC<sub>5</sub>, opioid<sub>κ</sub>, and pA<sub>2</sub> receptors (Compton et al., 1996; Shire et al., 1996).

Functional studies confirmed its potent (pA<sub>2</sub> 7.98–8.85) and selective CB1 receptor antagonistic activity. This compound readily displaced [<sup>3</sup>H]CP 55,940 from specific binding sites ( $K_i = 1.98$  nM) and has been shown to prevent cannabinoids from producing several of their typical effects, both in vitro and in vivo. Rimonabant potency as an antagonist has been shown by comparing its ability to attenuate WIN 55,212-2-induced inhibition of electrically evoked contractions of the mouse isolated vas deferens ( $K_d = 2.4$  nM) with that of WIN 56,098, bromopravadoline, and iodopravadoline. It was also an effective antagonist in vivo by suppressing the hypothermia elicited by WIN 55,212-2 and psychomotor effects in mice and rats (Perio et al., 1996) (Table 1).

Saturation binding experiments with membranes prepared from rat cerebellum have shown that radiolabeled rimonabant undergoes specific, rapid, saturable, high-affinity binding to a single class of sites. This specific binding is little affected by micromolar concentrations of a variety of noncannabinoid receptor ligands. However, it is readily attenuated

by the cannabinoids CP 55,940, WIN 55,212-2,  $\Delta^9$ -THC, 11-hydroxy- $\Delta^9$ -THC, and AEA. Interactions with rimonabant are competitive in nature for CP 55,940, WIN 55,212-2, and  $\Delta^9$ -THC, but noncompetitive for AEA, because the latter compound decreases both the affinity constant and the  $B_{max}$  of radiolabeled rimonabant (Rinaldi-Carmona et al., 1994; Petit et al., 1996). Shire et al. (1996) have carried out experiments to identify the domain(s) of the cannabinoid CB1 receptor responsible for the recognition and binding of rimonabant. Their approach was to transfect COS-3 cells with mutated CB1 receptors or with a range of different chimeric CB1/CB2 receptors. The results obtained suggest that the fourth and fifth transmembrane domains of the CB1 receptor are essential for high-affinity binding of rimonabant, whereas the extracellular loop between these two domains is unimportant.

In some experiments, rimonabant has been found to produce effects that are opposite in direction from those produced by cannabinoid receptor agonists. In particular, it can increase locomotor activity in mice (Compton et al., 1996), improve social short-term memory in rats and mice (Teranova et al., 1996), augment forskolin-induced stimulation of cyclic AMP production in cells transfected with CB1 (Felder et al., 1995), increase the amplitude of electrically evoked contractions of various isolated tissue preparations (Pertwee et al., 1996), and enhance electrically evoked release of acetylcholine from both rat hippocampal slices and the myenteric longitudinal muscle of guinea pig small intestine (Gifford and Ashby, 1996).

Increasing evidence suggests that rimonabant behaves also as an inverse agonist in some membrane preparations. Indeed, Bouaboula et al. (1997) found that Chinese hamster ovary cells, transfected with the CB1 receptor, displayed high constitutive activity of both MAPK and adenylate cyclase and this increase was inhibited by rimonabant. They also observed that guanosine 5'-O-(3-[<sup>35</sup>S]thio)triphosphate enhanced the binding of rimonabant, a feature usually described for inverse agonists. The issue on inverse agonistic properties of rimonabant has been reviewed thoroughly by Pertwee (2005).

The pharmacokinetic/pharmacodynamic profile of rimonabant, as expected by both preclinical and clinical studies showed that rimonabant is distributed widely in brown fat, it could reduce total energy intake and body weight gain in obese rats, and the most effective dose in reducing body weight in obese human subjects was 20 mg/day (Table 1).

### Effect of Rimonabant on Weight Loss and Cardiovascular Risk Factors

The assessment of the clinical efficacy of rimonabant as an antiobesity drug was carried out in multinational, randomized and placebo-controlled trials on patients who were overweight (body mass index higher than 27 kg/m<sup>2</sup>) or obese (body mass index  $\geq 30$  kg/m<sup>2</sup>) (Van Gaal et al., 2005; Pi-Sunyer et al., 2006). A cumulative weight loss and a significant change of waist circumference from the baseline were observed in patients receiving 20 mg/day of rimonabant. It is noteworthy that rimonabant also caused an increase of high-density lipoprotein (HDL) cholesterol levels and a reduction of triglycerides, fasting insulin, and insulin resistance derived from homeostasis model assessment, which was calcu-

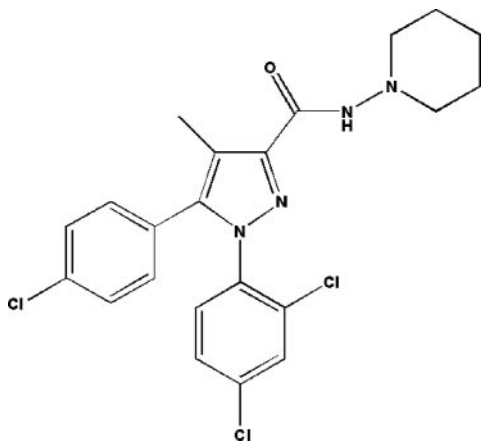


Fig. 1. Chemical structure of rimonabant.

lated by multiplying fasting insulin by fasting glucose and dividing by 22.5 (Pi-Sunyer et al., 2006).

After 1 year of rimonabant treatment (20 mg/day), the levels of HDL cholesterol, triglycerides, and the other above-mentioned parameters were twice those attributable to the concurrent weight loss alone as assessed by analysis of covariance (Pi-Sunyer et al., 2006). In addition, in patients who completed the study in the second year and received 20 mg of rimonabant, levels of triglycerides and fasting insulin declined rapidly from baseline, suggesting a direct pharmacological effect of rimonabant on glucose and lipid metabolism outside the weight loss achieved (Pi-Sunyer et al., 2006).

In another recent trial aimed to assess the effects of rimonabant in overweight patients with dyslipidemia (Désprés et al., 2005), a significant weight loss was reached in the majority of patients completing the 12 month study and receiving 20 mg/day of rimonabant. During the treatment, the levels of triglycerides significantly decreased, whereas HDL cholesterol increased compared with both placebo group and that treated with 5 mg/day rimonabant. Furthermore, the prevalence of metabolic syndrome, a collection of factors (abdominal adiposity, hypertriglyceridemia, low HDL cholesterol, hypertension, and fasting hyperglycemia) increasing the risk of type 2 diabetes and cardiovascular disease (Nesto, 2005), was reduced in the same subset of patients (Pi-Sunyer et al., 2006). Indeed, the population of patients matching the National Cholesterol Education Program's Adult Treatment Panel III (NCEP-ATPIII) criteria for the metabolic syndrome

(54% of the total) fell to the half of baseline after they received 20 mg of rimonabant (Désprés et al., 2005).

The overall observations strongly suggest that rimonabant could modulate positively risk factors for a number of obesity related comorbidities through weight loss-dependent and -independent pathways. In the sense of testing the clinical efficacy of the drug in new therapeutic strategies, we point out that future research should be focused on its potential pharmacological action in primary clinical outcome of the metabolic syndrome (e.g., cardiovascular disease, coronary heart disease, cerebrovascular disease, and peripheral artery disease, which have been associated with the high prevalence of the metabolic syndrome) (Nesto, 2005). It is noteworthy that the treatment with 20 mg of rimonabant for 1 year increases significantly the levels of plasma adiponectin compared with the placebo group (Désprés et al., 2005). This finding is not unexpected because an enhanced adiponectin serum level has been associated with weight reduction (Yang et al., 2001). Indeed, serum adiponectin levels are inversely correlated to obesity, metabolic syndrome, and type 2 diabetes (Gable et al., 2006); therefore, the observation that rimonabant improves the levels of either adiponectin or fasting insulin and induces favorable changes in insulin resistance derived from homeostasis model assessment, strongly suggests a possible pharmacological application of rimonabant in diabetes. Recently, it has been demonstrated that 20 mg/day of rimonabant can improve a number of cardiovascular

TABLE 1  
Pharmacodynamics and pharmacokinetics of rimonabant  
Data at <http://www.emea.eu.int>.

	Preclinical Studies	Clinical Studies
Primary and secondary pharmacodynamics	Potent and selective CB1 antagonistic activity in vitro ( $pA_{50}$ , 7.98–8.85) Reversion of WIN55212-2-induced hypothermia and psychomotor effects in mice and rats (either i.p. or p.o.; $ID_{50}$ from 0.2 to 1.7 mg/kg) Long-lasting and selective inhibition in sucrose drinking (75%) in models for obesity (1 and 3 mg/kg, p.o.) Reduced intake of the preferred high-fat diet (37%), total energy intake in obese rats (by 30%) and body weight gain (93%) (3 mg/kg/day; for 12 days, p.o.). Reduction of known risk factors associated with obesity, serum leptin (81%), insulin (78%), glucose (67%) (10 mg/kg/day) Reduction of nicotine self-administration at 0.3 mg/kg and cue-induced nicotine seeking at 1 mg/kg in the self-administration paradigm and at 3 mg/kg in the conditioned place preference test in several animal models (see text). Increase of the number of shocks (0.3 mg/kg) in the punished drinking test (3 mg/kg) and percentage of time spent into open arms at 10 mg/kg in the elevated plus maze test. Decrease of immobility time from 3 mg/kg, p.o. in the forced swimming test.	Obesity: reduction of hunger, daily caloric intake and mean body weight (7-day repeated doses of 20 mg) Antagonism of cannabis effects: inhibition of cannabis-induced effects [e.g., heart rate increase (90-mg single dose, 40-mg repeated doses)].
Pharmacokinetics:		
Adsorption	Oral bioavailability low to moderate (12% male rats, 46% female rats, 18% male macaques) Extensive first pass metabolism peak concentration after 1–3 h steady state pharmacokinetic: 1 week once-daily dosing.	Rapidly absorbed upon oral administration. Decreased absorption with increased dose; $C_{max}$ reached 2 h after 20-mg dose; low solubility and high permeability Steady state: 13 days in normal-weight volunteers, 39 days in obese patients
Distribution	Volume of distribution > total body water (11.5 l/kg rats, 24.4 l/kg macaques) Tissues with greatest uptake: liver, adrenals, brown fat, kidneys, lymph nodes; High binding (up to 100%) to plasma proteins.	Never administered i.v. Extensive distribution Very high binding to plasma proteins, mainly albumin (mean-99.9%)
Metabolism	Amidohydrolysis, oxidative metabolism on the piperidine moiety, glucuronidation of the acid, hydroxy phase 1 metabolism.	Metabolized by CYP3A4 and hepatic amidohydrolyase Metabolites inactive against human cloned CB1 receptors
Excretion	Long terminal half-life in rats (7.3 h) and macaques (20 h) and medium clearance (~1 l/h/kg) Excretion via bile in the feces (>70%), small amount in urine. Excreted in milk (rats).	Mean terminal half-life: 10 days, in normal-weight volunteers, 16 days, in obese patients. Clearance, 5 l/h. After a 20 mg/kg dose: 32% excretion as unchanged in feces; 3% in urine and 61% in feces over 312 h. Biliary excretion of metabolites



and metabolic risk factors in patients who are overweight or obese and have type 2 diabetes (Scheen et al., 2006).

To date, safety evaluation and recording of adverse events from all RIO studies have reported nausea, diarrhea, and upper respiratory tract infections in the first month of treatment. Serious adverse effects, such as psychiatric (depressed mood disorders), nervous system, and gastrointestinal disorders occur more frequently in patients treated with 20 mg/day compared with the patients in the placebo group during the first year (Déspres et al., 2005; Van Gaal et al., 2005; Pi-Sunyer et al., 2006). Further studies are required to assess the long-term effects of rimonabant, beneficial or adverse, beyond 2 years.

Although rimonabant has been shown to be a powerful agent for the treatment of obesity, as assessed by the mentioned clinical trials, its biological mechanism of action remains unclear. Key questions still have to be tackled: Is the weight loss induced by the CB1 receptor blocker rimonabant a consequence of an action to central level or is it the result of a control at peripheral energy metabolism? What signal transducing events are involved in weight loss?

In nongenetic-induced obese mice, body weight loss seems to consist of an early phase that depends on regulation of food intake and a second phase that is independent of food intake regulation in which weight loss is maintained probably through a sustained reduction of adiposity. Therefore, it is conceivable that rimonabant interferes with both the regulation of the expression and release of hypothalamic neuropeptides involved in the control of appetite and with peripheral metabolism. In agreement with this hypothesis, the treatment with rimonabant of mice lacking the cocaine-amphetamine-regulated transcript, an anorexigenic peptide, does not affect feeding behavior (Ravinet Trillou et al., 2003), whereas this compound inhibits starvation-induced hyperphagia in neuropeptide Y-deficient mice and reduced body weight in leptin knockout mice (Di Marzo et al., 2001). The reduction of body weight by rimonabant is likely to involve the modulation of more than one orexigenic pathway controlling food intake at central level. In this sense, recent data showing a functional colocalization of CB1 receptor and orexin 1 receptor, have demonstrated that the treatment with rimonabant of Chinese hamster ovary cells expressing both receptors, completely prevented orexin response, thus providing further insights that the weight loss achieved in vivo could be partly ascribed to the blockade of CB1/orexin 1 receptor cross-talk (Hilairet et al., 2003). On the other hand, the finding that the CB1 receptor-mediated lipogenic activity in primary murine adipocytes can be blocked by rimonabant (Cota et al., 2003) intriguingly supports the idea that this compound may also function at peripheral level by decreasing lipoprotein lipase activity. In line with this hypothesis, in obese Zucker (fa/fa) rats, rimonabant stimulates Acrp30 mRNA, an adipocytokine exclusively expressed by adipocytes and regulating fatty acid oxidation. This stimulation has been also found to occur within 30 min of rimonabant treatment of cultured mouse 3T3 F442A preadipocyte cells (Bensaid et al., 2003). The Acrp30 mRNA inhibition was accompanied by a reduced adipocyte proliferation and an enhanced cell maturation (Gary-Bobo et al., 2006). The effect is mediated by the inhibition of MAPK phosphorylation. It is noteworthy that rimonabant, working as an inverse agonist, is able to switch off MAPK activation from the insulin recep-

tor-tyrosine kinase and insulin-like growth factor receptors (Bouaboula et al., 1997). Moreover, endocannabinoid tonic activation of CB1 receptor in liver induces mRNA expression of the lipogenic transcription factor sterol regulatory element binding protein-1c, its target enzymes acetyl coenzyme-A carboxylase, and fatty acid synthase, also increasing the rate of fatty acid synthesis (Osei-Hyiaman et al., 2005). Blockage of CB1 receptor stimulation by rimonabant significantly reduces de novo fatty acid synthesis in mice, thus providing the evidence for the involvement of fatty acid biosynthetic pathway in the sustained reduction of body weight. The reported outcomes explain how rimonabant induces its effects at central and peripheral levels, even though it would be of interest to study whether other G-protein and/or orexigenic pathways could be responsible for the rimonabant-induced antiobesity effects and whether there are mechanisms of action independent from the interaction with the CB1 receptor.

### Rimonabant-Induced Effects on Drug Dependence

CB1 receptors are expressed at high levels in brain regions thought to play a key role in relapse-like behavior and conditioning processes. In these regions, CB1 receptors modulate the release of a variety of neurotransmitters (De Vries and Schoffelmeer, 2005) implicated in drug seeking behavior, smoking cessation, and alcohol addiction, suggesting that rimonabant could offer a novel approach for the treatment of behavior-related disorders (for review, see Beardsley and Thomas 2005; De Vries and Schoffelmeer, 2005).

**Smoking Cessation.** Pharmacological studies aimed to investigate the effect of rimonabant on motivational effects of nicotine in rats, demonstrated that a single administration of rimonabant (0.3 and 1.0 mg/kg) on two consecutive days reduced nicotine infusions and the presses of active lever from the first day of treatment (Cohen et al., 2002). Given the findings that rimonabant reduced responding for nicotine-associated cues, even after several months of nicotine abstinence, and antagonizes dopamine release caused by nicotine addiction, it is reasonable that the CB1 receptor antagonist can reduce both nicotine-seeking behavior and nicotine-reinforcing effects, probably by preventing nicotine-induced dopamine (DA) release in limbic dopaminergic areas during self-administration (Cohen et al., 2002, 2005). The action of rimonabant on both reducing nicotine craving and weight gain may be, at least in part, the result of a common mechanism involving a dopamine receptor-mediated process (Duarte et al., 2004). In this respect, studies of rimonabant treatment on conditioned response for nicotine- and sucrose-associated cues in a long-term extinction-reinstatement animal model demonstrated a strongly decrease cue-induced reinstatement of nicotine- and sucrose-seeking behavior (De Vries et al., 2005). The blockade of CB1 receptors attenuated reinstating responding for stimuli associated with nicotine infusion and sucrose delivery in a similar fashion. Preclinical studies seem to support this evidence: 1) rimonabant reduces nicotine self-administration, DA turnover in nucleus accumbens, and reinstatement of nicotine-seeking behavior (Fagerstrom and Balfour, 2006); 2) The Studies with Rimonabant And Tobacco Use (STRATUS-US), studying 787 smokers at 11 clinical trial sites in the United States, showed that 36% of patients treated with 20 mg/day rimonabant quit smoking,

whereas only 20.6% of patients treated with placebo and 20.2% of patients treated with 5 mg/day of rimonabant were successful (Anthenelli and Despres, 2004). Previous findings suggest that rimonabant may represent a novel drug for the treatment of tobacco dependence and may be effective not only as an aid for smoking cessation but also in the maintenance of abstinence.

**Ethanol Dependence.** A large body of research lines suggests functional interactions between the cannabinoid receptor CB1 and ethanol dependence; CB1 receptor is also involved in the mechanism mediating alcohol relapse (Maldonado et al., 2006). Rimonabant treatment induces a decrease in voluntary ethanol intake in alcohol-preferring rats (Colombo et al., 1998) and decreases ethanol consumption in mice (Arnone et al., 1997). Cippitelli and coworkers (2005) analyzed the effects of cannabinoid receptor blockade by rimonabant on alcohol self-administration and cue-induced relapse in Wistar and genetically selected Marchigian Sardinian alcohol preferring rats (msP) rats, in which the acquisition of ethanol self-administration was quicker than in wild-type strains. Rimonabant was found to reduce either ethanol self-administration or cue-induced relapse to ethanol self-administration and was more efficacious in the msP rats than in Wistar rats. It is noteworthy that strong differences in CB1 receptor mRNA levels between ethanol-naive msP and Wistar rats have been found, suggesting that the effects of rimonabant could be ascribable to the genetically determined different levels of CB1 receptor in brain regions responsible for ethanol dependence. This assumption is also supported by the finding that CB1<sup>-/-</sup> mice and rimonabant-treated mice consume significantly less alcohol than the corresponding untreated wild-type mice (Wang et al., 2003). However, a reduction of the reinforcing properties of ethanol in the self-administration paradigm has also been observed in animal models of ethanol seeking elicited by environmental stimuli (Economidou et al., 2006). Therefore, considering the rimonabant-mediated reduction of reward-related responding to ethanol, it could be of relevant interest to test this compound in preclinical studies, aimed at proving its pharmacotherapeutic properties in the treatment of ethanol dependence.

**Psychostimulants and Opioid Seeking.** Although rimonabant administration does not interfere with cocaine self-administration in monkeys (Tanda et al., 1997) and relapse induced by exposure to stress in rats, it reduced relapse to cocaine-seeking behavior produced by re-exposure to cocaine-associated cues (De Vries et al., 2001). Likewise, rimonabant blocked the relapse of methamphetamine-seeking behavior in rats also preventing the reinstatement of methamphetamine-seeking behavior, when administered before the cue phase of the test session (Anggadiredja et al., 2004).

Concerning opioids, several studies have revealed solid evidence for the existence of a functional interaction between cannabinoid and opioid system, suggesting that CB1 receptor may play an important role in the mechanism underlying relapse to heroin seeking (Tanda et al., 1997; Navarro et al., 2001). For the most part, in opiate self-administration and opiate-induced place preference in rats and mice, using heroin and morphine, acute administration of rimonabant (3.0 mg/kg) reduced heroin self-administration in Wistar rats and blocked heroin and morphine self-administration in mice.

Because rimonabant does not interact directly with opioid receptors, it could interact with dopaminergic system in the nucleus accumbens, attenuating extracellular dopamine release (Navarro et al., 2001). Furthermore, rimonabant attenuated (by about 50%) the reinstatement of heroin-seeking behavior caused by both a priming injection of heroin and a re-exposure to heroin-paired stimuli. In addition, rimonabant dose-dependently reduced responding for heroin in fixed and progressive ratio schedules of reinforcement (De Vries et al., 2003; Spano et al., 2004). These observations indicate that the selective CB1 receptor antagonist rimonabant might be used to attenuate both the reinforcing/motivational properties of heroin and the reinstatement of heroin seeking after prolonged withdrawal (Fattore et al., 2005).

### Rimonabant-Induced Effects on Neurodegenerative Disorders

Until endocannabinoid system was discovered, the role of CB1 receptor in the physiology and pathology of nervous system has received particular attention because of its selective and relatively high expression within the central nervous system. The CB1 receptor is localized mostly in the brain (Herkenham et al., 1990) and spinal cord (Herkenham et al., 1991a) and is expressed in the output nuclei of basal ganglia, in the substantia nigra pars reticulata and globus pallidus. Intermediate receptor levels have also been found in the cortex, hippocampus, thalamic nuclei, hypothalamus, and cerebellum (Herkenham et al., 1990, 1991a,b; Jansen et al., 1992; Thomas et al., 1992). The endogenous ligands of CB1 receptor are synthesized upon demand by neurons in response to depolarization (Freund et al., 2003); once released from postsynaptic neurons, they can function as retrograde synaptic messengers. They travel backward across synapses, activate CB1 on presynaptic axons, and inhibit neurotransmitter release (Szabo et al., 1998; Wilson and Nicoll, 2002). Because of these properties, the endocannabinoid system could offer new pharmacological targets to alleviate motor symptoms and supply neuroprotection in neurological disorders such as Parkinson's and Alzheimer's disease, Huntington's chorea, and multiple sclerosis. Despite the projectile findings on this issue, recently well reviewed (Fernandez Ruiz and Gonzales, 2005; Robson, 2005; Valverde et al., 2005; Walker and Hohmann, 2005; Pertwee, 2006), inconclusive results were reported on early study carried out with rimonabant. Rimonabant increased the frequency and duration of seizures in a rat model of viral encephalopathy (Borna disease virus rats) (Solbrig et al., 2005), whereas the hyperkinetic state (vertical activity) induced by L-DOPA was decreased by the subcutaneous injection of rimonabant in the reserpine-treated rat model of Parkinson's disease. On the other hand, discrepant results have been obtained about the effects of this compound on quinpirole-induced hyperactivity. Rimonabant administered alone has no evident effects on motor activity (Giuffrida et al., 1999; Segovia et al., 2003); when injected 1 h before quinpirole, however, it potentiates motor stimulation, thus suggesting a complex interaction between CB1 receptor, its agonists and dopamine receptors (Segovia et al., 2003). Moreover, rimonabant reverses the neuroprotective effect of CB1 agonists in primary neuronal cultures from spinal cords *in vitro* (Abood et al., 2001) and in animal models of both genetic and pharmacologically induced

Huntington's disease in vivo (Lastres-Becker et al., 2003; Centonze et al., 2005), also causing epileptic activity during development (Bernard et al., 2005). It is surprising that rimonabant was ineffective in producing hyperalgesia in rats and by itself is able to exert antinociceptive effect, probably by blocking the action of CB1 receptor stimulation on presynaptic GABA release (Naderi et al., 2005).

### Rimonabant Antiproliferative Effects

Because hyperplasia of adipose tissue is a crucial event for the development of obesity, the antiproliferative effect of rimonabant has been investigated on mouse preadipocytes. A reduced proliferation and an induced late maturation of adipocytes, without lipid droplet accumulation, mediated by an inhibition of basal and serum-induced p42/44 MAPK pathway was observed (Gary-Bobo et al., 2006). The MAPK pathway, strongly activated by a high-fat diet in white adipose tissue and required for the development of obesity (Bost et al., 2005), is inhibited by rimonabant through pertussis toxin-sensitive tyrosine kinase receptors, such as those for insulin or insulin-like growth factor 1, therefore displaying a negative intrinsic activity ascribable to inverse agonism (Bouaboula et al., 1997; Landsman et al., 1997).

The endocannabinoid system is implicated in the pathogenesis of chronic liver diseases associated with hepatic fibrosis (Mallat and Lotersztajn, 2006). CB1 receptors in particular are highly up-regulated in human cirrhotic specimens and in liver fibrogenic cells. Recent findings have emphasized the idea that CB1 receptor antagonism by rimonabant administration in mice counteracts the wound-healing response to acute liver injuries by decreasing the accumulation of hepatic myofibroblasts and the levels of the profibrogenic cytokine transforming growth factor  $\beta$ 1. The antiproliferative effect depends on CB1 receptor signaling, as revealed by the absence of antiproliferative effects in *Cnr1*<sup>-/-</sup> hepatic myofibroblasts. As previously observed in several cell systems, the molecular pathways mediating CB1 antagonism effects involve decreased phosphorylation of extracellular signal-regulated kinase MAPK and Akt, both in *Cnr1*<sup>-/-</sup> cells and in wild-type cells treated with rimonabant (Teixeira-Clerc et al., 2006).

Together, these findings provide evidence for an antifibrotic effect of rimonabant and suggest that rimonabant might represent a therapeutic tool for the treatment of some pathological liver conditions in humans.

### Rimonabant Antitumor Effects

The studies conducted from the late 1990s on the endocannabinoid system have provided strong evidence for a key role of the endocannabinoids in the control of cancer cell growth, invasion, and metastasis processes in a way dependent on CB receptor activation (for review, see Bifulco and Di Marzo, 2002; Bifulco et al., 2006, 2007).

The selective CB1 receptor antagonist rimonabant attenuates the antitumor effects of anandamide-related compounds or other cannabinoid agonists in thyroid, breast, and prostate cancers (Bifulco et al., 2001; Portella et al., 2003; Sarfaraz et al., 2005; Grimaldi et al., 2006); the effects are dependent on CB1 receptor activation. In other tumor types, such as glioma, rimonabant failed to revert the antiproliferative action

of cannabinoid agonists, whereas the selective CB2 antagonist SR144528 (Sanchez et al., 2001) or a combination of the CB1/CB2 antagonists can partially prevent this effect (Jacobsson et al., 2001). However, a 48-h preincubation with these antagonists seems to enhance the AEA-mediated cell death of glioma cells, suggesting a more complex mechanism of action (Maccarrone et al., 2000a).

Considering the antitumor properties of the cannabinoid receptor agonists, it could be expected that cannabinoid receptor antagonists, such as rimonabant, if used alone, would enhance proliferation of normal and malignant cells, leading to cancer. Some data excluded this possibility, reporting rather that not only agonists to cannabinoid receptors but also antagonists, used alone, are able to inhibit cancer growth (Bifulco et al., 2004) or induce apoptosis in cancer cells (Derocq et al., 1998; Powles et al., 2005).

Our group provided the first observation of a potential antitumor action in rimonabant in rat thyroid cancer cells (KiMol) in vitro and in thyroid tumor xenografts induced by KiMol injection in athymic mice. In this model, rimonabant was able to partially prevent the antitumor effect of the inhibitors of endocannabinoid degradation and of the anandamide metabolically stable analog (2-methylarachidonyl-2'-fluoro-ethylamide). However, rimonabant, when used alone, in the same model and at the same dose shown previously to counteract the 2-methylarachidonyl-2'-fluoro-ethylamide effect (0.7 mg/kg intratumoral, twice a week for two weeks), did not enhance tumor growth, exerting a small but significant antitumor effect on thyroid tumors, both in vitro and in vivo (Bifulco et al., 2004).

It is noteworthy that micromolar concentrations of rimonabant decreased viability of primary mantle lymphoma cells isolated from tumor biopsies of two patients after treatment with micromolar concentrations of rimonabant (Flygare et al., 2005). Moreover, rimonabant showed an additive negative effect as well on the viability of the mantle cell lymphoma cell line Rec-1 when combined with equipotent doses of AEA. Bifulco et al. (2004) and Flygare et al. (2005) supported the evidence of the antitumor action of rimonabant, but they did not investigate or provide a molecular mechanism of action. They proposed that the observed effects could be ascribed to: 1) a tonic antiproliferative action mediated by the local endocannabinoids through mechanisms independent from CB1 receptor, particularly when CB1 receptors are blocked by the antagonist rimonabant; and 2) the inverse agonist properties of rimonabant on the receptor. These possibilities could explain the paradox whereby both CB1 agonists and antagonists display antitumor activity.

We have reported that rimonabant exerts antitumor effects on breast cancer in vitro and in a mouse model in vivo, providing for the first time a new mechanism of action for this drug (Sarnataro et al., 2006). Rimonabant, at nanomolar concentrations, is able to inhibit human breast cancer cell proliferation; it is more effective in highly invasive metastatic MDA-MB-231 cells than in less invasive T47D and MCF-7 cells, depending on both the presence and the different expression levels of the CB1 receptor. The antiproliferative effect is characterized by a G<sub>1</sub>/S phase cell cycle arrest, without induction of apoptosis. The in vitro observed effect has also been confirmed in vivo: after 2 weeks of treatment, rimonabant reduces the volume of xenograft tumors induced by MDA-MB-231 injection in mice. The molecular mecha-



nism at the basis of rimonabant function implicates an inhibition of p42/44 MAPK phosphorylation and requires lipid rafts/caveolae integrity. This suggests that rimonabant's effects on cell proliferation and signaling requires the presence of CB1 receptor in lipid rafts (Sarnataro et al., 2006).

### Rimonabant-Induced Effects on Fertility

During the last few years, accumulating evidence has indicated that the endocannabinoid system may play an important role in modulating reproductive system functions and fertility. Some reports have underlined the presence of both CB1 receptor subtype in human sperm (Rossato et al., 2005) and significant concentration of endocannabinoids in female and male genital tract fluids (Schuel et al., 2002). This finding suggests that the control of the endogenous tone of endocannabinoids and its interaction with the CB receptors are checkpoints in reproduction (for review, see Maccarrone and Finazzi-Agrò, 2004). However, CB1 receptor activation by AEA is responsible for a reduced sperm motility and inhibition of capacitation-induced acrosome reaction in human sperm specimens. The CB1-selective antagonist rimonabant is able to block the negative effects of AEA on motility of sperm without compromising sperm viability or motility per se (Rossato et al., 2005). Moreover, Melis and coworkers (2006) reported that rimonabant was able to induce penile erection in male rats when injected into the paraventricular nucleus of hypothalamus. This effect was associated with an increase of glutamic acid leading to the activation of neuronal and nitric-oxide synthase in oxytocinergic neurons mediating penile erection (Succu et al., 2006). However, it is possible that rimonabant-induced penile erection also implies an increase in dopaminergic neurotransmission (da Silva et al., 2003).

Few data on the effects of rimonabant on fertility are available, whereas a large body of the recent literature has been focused on the interaction and possible regulation of reproductive processes by endocannabinoid system. Endocannabinoids are involved in implantation (attachment and outgrowth of blastocysts), pregnancy (myometrial contractility) (Liu et al., 2002; Denny et al., 2004), and miscarriage (Maccarrone et al., 2000b). Therefore, it is possible to speculate that not only a decreasing AEA concentration in human reproductive tract secretions but also the administering of rimonabant may represent therapeutic tools in pathological situations such as recurrent abortions characterized by increased levels of AEA.

### Other Effects of Rimonabant

The promising results obtained in several experimental model systems, proposing rimonabant as a potential therapeutic tool for the treatment of several pathological conditions, have recently promoted investigations to ascertain the potential benefit effects of this compound, mainly as a CB1 receptor antagonist, in other disorders affecting the central nervous system, the immune system, and the circulatory system. In this sense, the high concentration of cannabinoid CB1 receptors expressed in hippocampus suggests that the cannabinoid neurochemical system may play a role in learning and memory processes (Takahashi et al., 2005). Some evidence supports the idea that the natural and synthetic

cannabinoids impair cognitive processes in humans, nonhuman primates, and rodents (Braida and Sala, 2000) and seem to inhibit hippocampal extracellular acetylcholine release (Terranova et al., 1996). Rimonabant reverses many of the biochemical physiological and behavioral effects of cannabinoid receptor agonists; e.g., it attenuates the memory impairment produced by AEA and THC (Mallet and Beninger, 1998; Mishima et al., 2001). Rimonabant, which per se does not influence memory processes at the dose of 0.5 mg/kg, completely antagonizes the impairment produced by the synthetic cannabinoid CP 55,940 (Braida and Sala, 2000).

Finally, blockade of CB1 receptor by rimonabant improves amnesia induced by the  $\beta$ -amyloid fragment in mice, suggesting that endogenous cannabinoids may be involved in cognitive impairment induced by these fragments. The injection of rimonabant alone does not cause any significant change in the capacity of mice to retain passive avoidance responses, and its effect may be observed only when CB1 are activated by their antagonists (Mazzola et al., 2003). From these few available data, it is not yet possible to support a potential therapeutic application of rimonabant in memory impairment or to propose a mechanism of action; therefore, further research is needed in this direction. Because CB1 receptor mRNA has been detected outside the brain in many other tissues, including immune system cells (Klein et al., 2003), recent articles on the role of endocannabinoids in the modulation of the immune system have led researchers to consider the therapeutic potential of rimonabant in the inflammation process.

Croci et al. (2003) proposed that rimonabant may interfere with immune-inflammatory pathogenic mechanisms such as that underlying indomethacin's ulcerogenic action. They observed that oral administration of rimonabant is able to dose-dependently prevent indomethacin-induced small intestinal ulcers in rats. This effect was associated with a higher inhibition of  $\text{TNF}\alpha$  levels and myeloperoxidase activity compared with CB2 receptor antagonist SR144528. Rimonabant produced similar inhibitory effects in CB1 receptor knockout mice, suggesting that its antiulcerogenic action does not rely on CB1 antagonism. However, in the same CB1 knockout mice, rimonabant failed to counteract the increase of LPS-induced  $\text{TNF}\alpha$  plasma levels that is CB1 receptor-dependent. This observation suggests that rimonabant could probably act with distinct mechanism of actions, modulating the inflammatory process in different ways. The inhibition of  $\text{TNF}\alpha$  levels by rimonabant is of particular interest, because an increase in plasma levels of this cytokine has been found in patients who are obese, and it could be involved in the regulation of glucose transport and insulin sensitivity (Hube and Hauner, 1999). In supporting the rimonabant anti-inflammatory action, its systemic administration could improve rat survival and endotoxin LPS-induced hypotension (Varga et al., 1998). It is noteworthy that the inhibition of LPS-induced hypotension by rimonabant does not depend on the presence of CB1 receptor, because rimonabant induces similar effects in CB1-deficient mice (Bátkai et al., 2004). Furthermore, other findings also pointed out that rimonabant was able to raise blood pressure, perhaps by counteracting the increased expression of CB1 receptor (Bátkai et al., 2001). On the bases of the previous findings, it seems clear that the role of cannabinoid system and the complex action of rimonabant on the circulatory system and its patho-

TABLE 2  
Overview of rimonabant effects

Pathological Conditions	Preclinical Studies		Clinical Studies	Effects	References
	In Vitro Models	In Vivo Models			
Obesity, metabolic syndrome, and associated cardiovascular diseases	Mouse preadipocytes	Nongenetic-obese mice, C57BL/6J and CB1 <sup>-/-</sup> mice		Inhibition of cell proliferation and MAPK activity; induction of adiponectin and GAPDH mRNA	Gary-Bobo et al., 2006
Neurodegenerative disorders	Primary neurons	BDV rats, reserpine treated rats, quinpirole-treated rats, R6/2 Huntington's disease mice, malonate-treated rat, Wistar rats, Wistar, Long-Evans, Lister Hooded rats; Wistar, Wistar/msP rats, Wistar rats, Sprague-Dawley rats	RIO Lipids, RIO Europe, RIO North America, RIO diabetes	Body weight loss and early reduction of food intake; weight loss maintained with normal diet Reduction of de novo fatty acid synthesis in mice Weight loss and reduced waist circumference; increased HDL levels and reduction of triglycerides, fasting insulin and insulin resistance; weight loss reduction, improvement of glucose control and metabolic parameters in type 2 diabetes Blockade of neuroprotective effects of CB1 receptor agonists Increase of convulsive phenomena; reduction of hyperkinetic state; potentiation of hyperkinesias; blockade of neuroprotective effects of CB1 receptor agonists; inhibition of neuroprotection; epileptic activity during development; analgesic properties Cocaine, attenuation of drug and cue-induced reinstatement; heroin, suppression of drug-induced reinstatement and attenuation of cue-induced reinstatement; methamphetamine, blockade of reinstatement; alcohol, reduction of cue induced reinstatement; attenuation of cue induced reinstatement, reduction of conditioned behavior after withdrawal 36% patients quit smoking Inhibition of thyroid tumor growth; decreased viability; additive effect with anandamide; antiproliferative effect with G <sub>1</sub> /S phase cell cycle arrest, linked to lipid rafts/caveolae localization of CB1 receptor signaling; reduced volume of xenograft tumors in mice Antagonizing the negative effects of increased levels of anandamide on sperm motility and acrosome reaction. Enhancement of penile erection Reduction of wound-healing response to acute liver injury; inhibited progression of fibrosis; growth inhibition of hepatic myofibroblasts Prevention of indomethacin-induced ulcers in rats and mice	Di Marzo et al., 2001; Arnone et al., 2003; Bensaid et al., 2003; Cota et al., 2003 Osei-Hyiaman et al., 2005 Despres et al., 2005; Van Gaal et al., 2005; Pi-Sunyer et al., 2006; Scheen et al., 2006; Abood et al., 2001 Giuffrida et al., 1999; Lastres-Beker et al., 2003; Segovia et al., 2003; Bernard et al., 2005; Centonze et al., 2005; Naderi et al., 2005; Solbrig et al., 2005 De Vries et al., 2001, 2003, 2005; Anggareddia et al., 2004; Athanelli and Despres, 2004; Spano et al., 2004; Cippitelli et al., 2005; Cohen et al., 2005; Economidou et al., 2006 Bifulco et al., 2004; Flygare et al., 2005; Sarnataro et al., 2006 Rossato et al., 2005 Da Silva et al., 2003; Melis et al., 2006 Teixeira-Clerc et al., 2006 Croci et al., 2003
Drug seeking (psychostimulants and opioids), alcohol dependence, Nicotine dependence			STRATUS-US		
Cancer	Rat thyroid cancer KiMol cells; human mantle cell lymphoma (MCL); Human breast cancer cells MDA-MB-231	Thyroid cancer xenografts in athymic mice Breast cancer xenografts in athymic mice			
Infertility	Human sperm	Sprague-Dawley rats			
Liver fibrosis	Human and murine hepatic myofibroblasts	CD1, C57BL/6J, Chr1 <sup>-/-</sup> mice			
Chronic inflammatory diseases: ulcer		CrI:CD BR rats C57BL/6J and CB1 <sup>-/-</sup> mice			



logic conditions needs further insights, to better clarify the molecular mechanisms and the signaling pathways evoked by rimonabant in determining such CB1-dependent and -independent effects.

## Conclusions and Perspectives

Collected results clearly show that rimonabant can have a plethora of pharmacological effects in a number of physiopathological conditions (Table 2). Because of rimonabant's selectivity for the cannabinoid CB1 receptor, the effects are mainly ascribable to its antagonistic properties, even though some evidence for its inverse agonistic action has also been provided (Bouaboula et al., 1997; Landsman et al., 1997; Navarro et al., 2001). Rimonabant represents a promising therapeutic tool in the treatment of obesity, as evidenced by clinical trials, and weight loss is achieved probably via central and peripheral mechanisms. Rimonabant is able to centrally target food intake regulation, acting on neurotransmitter release. The blockage of the CB1 receptor could result in a reduction of dopamine release that has been found to be enhanced in corticolimbic structures as a consequence of the rewarding effect of palatable food (Spanagel and Weiss, 1999). Furthermore, rimonabant reduced obesity in leptin (ob/ob) and leptin receptor (db/db) knockout mice (Di Marzo et al., 2001) and blocked CB1/OX1 receptor cross-talk (Hilairet et al., 2003), suggesting that it could exert a negative effect on genetically obese animals with an altered leptin neuroendocrine pathway and inhibit the CB1-mediated tonic orexigenic effect caused by increased levels of endocannabinoids. The modulation of nervous system functions is also at the basis of its pharmacological action on ethanol and sucrose consumption, drug-seeking behavior, and nicotine addiction. Above all, rimonabant may operate by preventing drug-induced DA release and turnover in dopaminergic areas, leading to an attenuation of either ethanol reward-related response or to nicotine-, psychostimulant-, and opioid-related relapse. However, by blocking CB1 receptor, rimonabant can act on GABAergic neurons stimulating GABA release (Naderi, 2005). This effect results in an induction of convulsive phenomena and epileptic activity in animal models of en-

cephalopathy and Parkinson's disease (Bernard et al., 2005; Solbrig et al., 2005). These observations, together with the finding that rimonabant prevents the benefit effects of CB1 receptor agonists in genetic and pharmacologically induced Huntington's disease (Lastres-Becker et al., 2003; Centonze et al., 2005), suggest that the drug is not suited to the treatment of neurodegenerative diseases and motor-related disorders. We should point out that rimonabant, used alone, does not induce an alteration of motor activity in normal rats; therefore, we could hypothesize that its adverse effects on nervous system functions could be limited to an already established pathological condition. On the other hand, this issue needs further evaluation to better clarify the exact role and physiopathological consequences of rimonabant-regulated GABA and DA release.

The molecular aspects at the basis of rimonabant-induced effects are not fully understood but seem to be related to the inhibition of the key steps of the CB1 signaling pathway. The molecular mechanism of action involves the inhibition of MAPK signaling. This pathway is activated by high-fat diet in white adipose tissue and is required for the development of obesity (Bost et al., 2005). Rimonabant, in agreement with its inverse agonistic properties, is able to inhibit CB1 receptor and switch off MAPK activation from the insulin receptor-tyrosine kinase and insulin-like growth factor receptors (Bouaboula et al., 1997). It is possible that this mechanism could be implicated also in the modulation of cardiovascular risk factors through a weight reduction-independent pathway, but how this occurs and the other biological pathways regulated by rimonabant and involved in glucose metabolism remain to be established. MAPK inhibition usually correlates with antiproliferative effects in both normal and cancer cells (Hou et al., 2004; Stepulak et al., 2005; Yoon and Seger, 2006) and the treatment of adipocytes and hepatic myofibroblasts with rimonabant strongly reduces proliferation through this pathway. More recently, we have provided evidence for an antiproliferative effect of rimonabant in thyroid and breast cancer cells (Bifulco et al., 2004; Sarnataro et al., 2006). In the last cellular model, we found that rimonabant strongly reduces cell growth by perturbing raft/caveolae integrity and

TABLE 3  
CB1 antagonists in clinical development  
Data at <http://www.clinicaltrials.gov>

Drug	Manufacturer	Clinical Phase	Study Type
Rimonabant	Sanofi-Aventis	III	Reducing the risk of major cardiovascular events in abdominally obese patients Effects on abdominal obese patients with dyslipidemia Amount and the activity of visceral fat in abdominally obese patients with metabolic syndrome Effects on abdominally obese patients with impaired fasting blood glucose with or without other comorbidities Effect on high density lipoprotein kinetics in patients with abdominal obesity and additional cardiometabolic risk factors
CP-945,598	Pfizer	II III	Reduction of voluntary ethanol drinking 2-year weight loss efficacy and safety in obese subjects Long-term study on weight loss and safety in obese subjects Obesity in overweight patients with type 2 diabetes
MK-0364	Merck	II	Weight maintenance in obese subjects Weight loss in obese and overweight subjects
Ave-1625	Sanofi-Aventis	II	Effects on abdominally obese patients with atherogenic dyslipidemia
SR147778	Sanofi-Aventis	II	Efficacy and safety in obese subjects
SLV-319	Solvay Pharmaceuticals Bristol-Myers Squibb	Moved to Phase II	

excluding CB1R from lipid rafts. We found that rimonabant's inhibitory effect on extracellular signal-regulated kinase 1/2 in the highly invasive and metastatic MDA-MB-231 breast cancer cells requires lipid raft integrity, thus suggesting that the role of lipid rafts in the receptor-dependent signaling would be to make favorable the CB1R-ligand encounter and the activation of CB1-dependent signaling (Sarnataro et al., 2006). Moreover, rimonabant causes a down-regulation of both the Acrp30 protein, sterol regulatory element binding protein-1c, and fatty acid synthase enzyme. At the same time, it is able to induce an up-regulation of adiponectin and glyceraldehyde 3-phosphate dehydrogenase, markers of adipose tissue functions, finally causing a reduction of adipocyte cell proliferation, a stimulation of fatty acid oxidation (Bensaid et al., 2003) or, alternatively, an inhibition of de novo fatty acid synthesis (Osei-Hyiaman et al., 2005).

Additional targets for the pharmacological effects of rimonabant include reproduction system functions. The role of the endocannabinoid system in reproduction and fertility has been reported (Dennedy et al., 2004; Maccarrone et al., 2004) but few available data demonstrated an increase in the number of penile erections in animal models that were probably due to an activation of dopaminergic and oxytocinergic neurotransmission mediated by rimonabant (da Silva et al., 2003; Melis et al., 2006; Succu et al., 2006). Therefore, it could be of great interest to intensify this issue, taking into account that the reproductive cascade of hormones and their regulation is tightly associated with energy metabolism and thus with the leptin pathway (Chehab, 2000). Finally, rimonabant seems to exhibit some beneficial effects in indomethacin-induced intestinal ulcer in rats, the effect being associated to a significant reduction of TNF $\alpha$  levels and myeloperoxidase activity (Croci et al., 2003), but these data on the potential application of rimonabant in inflammatory process, such as the results on a potential role of rimonabant in the treatment of memory impairment, are still quite scanty.

In light of the public health implications of the obesity pandemic, CB1 blockade strategy aimed to treat obesity and related disorders has encouraged several pharmaceutical companies to develop new and more selective CB1 antagonists, some of which are already in clinical trials (Table 3). At the moment, available data are not exhaustive to state the advantage of rimonabant competitor compounds; indeed, reported side effects are comparable and there are no data on long-term safety and efficacy. Moreover, in the pharmacotherapy of obesity, it would be necessary to take into account clinical and genetic parameters. We recently demonstrated a strong association between a polymorphic variant of CB1 receptor and glycemia and triglyceride concentration in patients who are overweight or obese (Gazzerro et al., 2007), and polymorphic variants in the codifying or promoting regions of CB1 receptor have been also associated to mood disorders and predisposition to depression (Barrero et al., 2005). Therefore, a subselection of patients' eligibility based on polymorphic CB1 receptor variants could influence the efficacy of the treatment and the incidence of side effects. Finally, rimonabant, showing antiobesity, anti-inflammatory, and antitumor properties, could be a preferential choice in breast cancer patients treated with chemotherapy, because excess adiposity is linked to risk of postmenopausal breast cancer, and the weight gain after chemotherapy is

linked with higher frequency of recurrent breast cancer (Harvie et al., 2005). Several authors proposed that local production of adipokines and inflammatory cytokines by adipocytes within the stroma surrounding breast epithelial cells may be directly linked to the growth of breast cancer (Manabe et al., 2003). Taken together, this evidence suggests that rimonabant could limit secretions by adipose tissue and improve the recurrence control in patients who have breast cancer and are obese and overweight.

In conclusion, we foresee other potential applications of rimonabant in related and nonrelated obesity pathologic conditions. On the bases of the pleiotropic effects described here, it represents a promise beyond its antiobesity action. Further studies will improve our understanding of the mechanisms of several diseases and will clarify the potential clinical impact of rimonabant "pleiotropic effects."

## References

- Abood ME, Rizvi G, Sallapudi N, McAllister S (2001) Activation of the CB1 cannabinoid receptor protects cultured mouse spinal neurons against excitotoxicity. *Neurosci Lett* **309**:197–201.
- Anggaredjaja K, Nakamichi M, Hiranita T, Tanaka H, Shoyama Y, Watanabe S, and Yamamoto T (2004) Endocannabinoid system modulates relapse to methamphetamine seeking: possible mediation by the arachidonic acid cascade. *Neuropsychopharmacology* **29**:1470–1478.
- Anthenelli RM and D  spret JP (2004) Effect of Rimonabant in the reduction of major cardiovascular risk factors. Results from the STRATUS-US trial (smoking cessation in smokers motivated to quit). American College of Cardiology 53rd Annual Scientific Session; 2004 Mar 7–10; New Orleans, LA.
- Arnone M, Maruani J, Chaperon F, Thiebaut MH, Poncelet M, Soubri   P, Le Fur G (1997) Selective inhibition of sucrose and ethanol intake by SR 141716, an antagonist of central cannabinoid (CB1) receptors. *Psychopharmacology* **132**:104–106.
- Barrero FJ, Ampuero I, Morales B, Vives F, de Dios Luna Del Castillo J, Hoenicka J, Garcia Yebenes J (2005) Depression in Parkinson's disease is related to a genetic polymorphism of the cannabinoid receptor gene (CNR1). *Pharmacogenomics J* **5**:135–141.
- B  tkai S, Zolt  n J, Wagner JA, Goparaju SK, Varga K, Liu J, Wang L, Mirshahi F, Khanolkar AD, Makriyannis A, et al. (2001) Endocannabinoids acting at vascular CB1 receptors mediate the vasodilated state in advanced cirrhosis. *Nat Med* **7**:827–832.
- B  tkai S, Pacher P, J  rai Z, Wagner JA, and Kunos G (2004) Cannabinoid antagonist SR-141716 inhibits endotoxic hypotension by a cardiac mechanism not involving CB1 or CB2 receptors. *Am J Physiol* **287**:H595–H600.
- Beardsley PM and Thomas BF (2005) Current evidence supporting a role of cannabinoid CB1 receptor (CB1R) antagonists as potential pharmacotherapies for drug abuse disorders. *Behav Pharmacol* **16**:275–296.
- Bensaid M, Gary-Boob M, Esclangon A, Maffrand JP, Le Fur G, Oury-Donat F, and Soubri   P (2003) The cannabinoid CB1 receptor antagonist SR141716 increases Acrp30 mRNA expression in Adipose tissue of obese fa/fa rats and in cultured adipocyte cells. *Mol Pharmacol* **63**:908–914.
- Bernard C, Milh M, Morozov YM, Ben-Ari Y, Freund TF, and Gozlan H (2005) Altering cannabinoid signaling during development disrupts neuronal activity. *Proc Natl Acad Sci USA* **102**:9388–9393.
- Bifulco M, Di Marzo V (2002) Targeting the endocannabinoid system in cancer therapy: a call for further research. *Nat Med* **8**:547–550.
- Bifulco M, Gazzerro P, Laezza C, and Pentimalli F (2007) Endocannabinoids as emerging suppressors of angiogenesis and tumor invasion. *Oncol Rep* **17**:813–816.
- Bifulco M, Laezza C, Portella G, Vitale M, Orlando G, De Petrocellis L, Di Marzo V (2001) Control by the endogenous cannabinoid system of ras oncogene-dependent tumor growth. *FASEB J* **15**:2745–2747.
- Bifulco M, Laezza C, Pisanti S, and Gazzerro P (2006) Cannabinoids and cancer: pros and cons of an antitumor strategy. *Br J Pharmacol* **148**:123–135.
- Bifulco M, Laezza C, Valenti M, Ligresti A, Portella G, Di Marzo V (2004) A new strategy to block tumor growth by inhibiting endocannabinoid inactivation. *FASEB J* **18**:1606–1608.
- Bost F, Aouadi M, Caron L, Even P, Belmonte N, Prot M, Dani C, Hofman P, Pages G, Pouyssegur J, et al. (2005) The extracellular signal-regulated kinase isoform ERK1 is specifically required for in vitro and in vivo adipogenesis. *Diabetes* **54**:402–411.
- Bouaboula M, Perrachon S, Milligan L, Canat X, Rinaldi-Carmona M, Portier M, Barth F, Calandra B, Pecceu F, Lupker J, et al. (1997) A selective inverse agonist for central cannabinoid receptor inhibits mitogen-activated protein kinase activation stimulated by insulin or insulin-like growth factor 1. Evidence for a new model of receptor/ligand interactions. *J Biol Chem* **272**:22330–22339.
- Braida D and Sala M (2000) Cannabinoid-induced working memory impairment is reversed by a second generation cholinesterase inhibitor in rats. *Neuroreport* **11**:2025–2029.
- Chehab FF (2000) Leptin as a regulator of adipose mass and reproduction. *Trends Pharmacol Sci* **21**:309–313.
- Centonze D, Rossi S, Prosperetti C, Tschertner A, Bernardi G, Maccarrone M, and Calabresi P (2005) Abnormal sensitivity to cannabinoid receptor stimulation might contribute to altered gamma-aminobutyric acid transmission in the striatum of R6/2 Huntington's disease mice. *Biol Psychiatry* **57**:1583–1589.



- Cippitelli A, Bilbao A, Hansson AC, del Arco I, Sommer W, Heilig M, Massi M, Bermudez-Silva FJ, Navarro M, Ciccocioppo R, et al. (2005) Cannabinoid CB1 receptor antagonism reduces conditioned reinstatement of ethanol-seeking behavior in rats. *Eur J Neurosci* **21**:2243–2251.
- Cohen C, Perrault G, Griebel G, and Soubrié P (2005) Nicotine-associated cues maintain nicotine-seeking behavior in rats several weeks after nicotine withdrawal: reversal by the cannabinoid (CB<sub>1</sub>) receptor antagonist, rimonabant (SR141716). *Neuropsychopharmacology* **30**:145–155.
- Cohen C, Perrault G, Voltz C, Steinberg R, and Soubrié P (2002) SR141716, a central cannabinoid (CB<sub>1</sub>) receptor antagonist, blocks the motivational and dopamine-releasing effects of nicotine in rats. *Behav Pharmacol* **13**:451–463.
- Colombo G, Agabio R, Fa M, Guano L, Lobina C, Loche A, Reali R, and Gessa GL (1998) Reduction of voluntary ethanol intake in ethanol-preferring sP rats by the cannabinoid antagonist SR141716. *Alcohol* **33**:126–130.
- Compton DR, Aceto MD, Lowe J, and Martin BR (1996) In vivo characterization of a specific cannabinoid receptor antagonist (SR141716A): inhibition of As-tetrahydrocannabinol-induced responses and apparent agonist activity. *J Pharmacol Exp Ther* **277**:586–594.
- Cota D, Marsicano G, Tschöp M, Grübler Y, Flachskamm C, Schubert M, Auer D, Yassouridis A, Thöne-Reineke C, Ortmann S, et al. (2003) The endogenous cannabinoid system affects energy balance via central orexigenic drive and peripheral lipogenesis. *J Clin Invest* **112**:423–431.
- Croci T, Landi M, Galzin AM, and Marini P (2003) Role of cannabinoid CB1 receptors and tumor necrosis factor- $\alpha$  in the gut and systemic anti-inflammatory activity of SR141716 (Rimonabant) in rodents. *Br J Pharmacol* **140**:115–122.
- da Silva GE, Fernandes MS, and Takahashi RN (2003) Potentiation of penile erection and yawning responses to apomorphine by cannabinoid receptor antagonist in rats. *Neurosci Lett* **349**:49–52.
- Denney MC, Friel AM, Houlihan DD, Broderick VM, Smith T, and Morrison JJ (2004) Cannabinoids and the human uterus during pregnancy. *Am J Obstet Gynecol* **190**:2–9.
- De Petrocellis L, Cascio MG, Di Marzo V (2004) The cannabinoid system: a general view and the latest additions. *Br J Pharmacol* **141**:765–774.
- Derooc JM, Bouaboula M, Marchand J, Rinaldi-Carmona M, Segui M, and Casellas P (1998) The endogenous cannabinoid anandamide is a lipid messenger activating cell growth via a cannabinoid receptor-independent pathway in hematopoietic cell lines. *FEBS Lett* **425**:419–425.
- Désprés JP, Golay A, Sjöström L, Rimonabant in Obesity-Lipids Study Group (2005) Effects of rimonabant on metabolic risk factors in overweight patients with dyslipidemia. *N Engl J Med* **353**:2121–2134.
- De Vries TJ, De Vries W, Janssen MC, and Schoffeleer AN (2005) Suppression of conditioned nicotine and sucrose seeking by the cannabinoid-1 receptor antagonist SR141716A. *Behav Brain Res* **161**:164–168.
- De Vries TJ, Homberg JR, Binnekade R, Raaso H, and Schoffeleer AN (2003) Cannabinoid modulation of the reinforcing and motivational properties of heroin and heroin-associated cues in rats. *Psychopharmacology (Berl)* **168**:164–169.
- De Vries TJ and Schoffeleer AN (2005) Cannabinoid CB1 receptors control conditioned drug seeking. *Trends Pharmacol Sci* **26**:420–426.
- De Vries TJ, Shaham Y, Homberg JR, Crombag H, Shuurman K, Dieben J, Vanderschuren LJ, and Schoffeleer AN (2001) A cannabinoid mechanism in relapse to cocaine seeking. *Nat Med* **7**:1151–1154.
- Di Marzo V, Bifulco M, De Petrocellis L (2004) The endocannabinoid system and its therapeutic exploitation. *Nat Rev Drug Discov* **3**:771–783.
- Di Marzo V, Goparaju SK, Wang L, Liu J, Bátkai S, Jári Z, Fezza F, Miura GI, Palmiter RD, Sugiura T, et al. (2001) Leptin-regulated endocannabinoids are involved in maintaining food intake. *Nature (Lond)* **410**:822–825.
- Duarte C, Alonso R, Bichet N, Cohen C, Soubrié P, and Thiébot ME (2004) Blockade by the cannabinoid CB1 receptor antagonist, rimonabant (SR141716), of the potentiation by quinelorane of food primed reinstatement of food-seeking behavior. *Neuropsychopharmacology* **29**:911–920.
- Economidou D, Mattioli I, Cifani C, Perfumi M, Massi M, Cuomo V, Trabace L, and Ciccocioppo R (2006) Effect of the cannabinoid CB1 receptor antagonist SR141716A on ethanol self-administration and ethanol-seeking behaviour in rats. *Psychopharmacology* **183**:394–403.
- Fagerstrom K, and Balfour DJ (2006) Neuropharmacology and potential efficacy of new treatments for tobacco dependence. *Expert Opin Investig Drugs* **15**:107–116.
- Fattore L, Spano S, Cossu G, Deiana S, Fadda P, and Fratta W (2005) Cannabinoid CB1 antagonist SR 141716A attenuates reinstatement of heroin self-administration in heroin-abstinent rats. *Neuropharmacology* **48**:1097–1104.
- Felder CC, Joyce KE, Briley EM, Mansouri J, Mackie K, Blond O, Lai Y, Ma AL, and Mitchell RL (1995) Comparison of the pharmacology and signal transduction of the human cannabinoid CB<sub>1</sub> and CB<sub>2</sub> receptors. *Mol Pharmacol* **48**:443–450.
- Fernandez-Ruiz J and Gonzales S (2005) Cannabinoid control of motor function at the basal ganglia. *Handb Exp Pharmacol* **168**:479–507.
- Flygare J, Gustafsson K, Kimby E, Christensson B, and Sander B (2005) Cannabinoid receptor ligands mediate growth inhibition and cell death in mantle cell lymphoma. *FEBS Lett* **579**:6885–6889.
- Freund TF, Katona I, and Piomelli D (2003) Role of endogenous cannabinoids in synaptic signaling. *Physiol Rev* **83**:1017–1066.
- Gable DR, Hurel SJ, and Humphries SE (2006) Adiponectin and its gene variants as risk factors for insulin resistance, the metabolic syndrome and cardiovascular disease. *Atherosclerosis* **188**:231–244.
- Gary-Bobo M, Elachouri G, Scatton B, Le Fur G, Oury-Donat F, and Bensaïd M (2006) The cannabinoid CB1 receptor antagonist rimonabant (SR141716) inhibits cell proliferation and increases markers of adipocyte maturation in cultured mouse 3T3 F442A preadipocytes. *Mol Pharmacol* **69**:471–478.
- Gazzerro P, Caruso MG, Notarnicola M, Misciagna G, Guerra V, Laezza C, Bifulco M (2007) Association between Cannabinoid type-1 receptor polymorphism and body mass index in a Southern Italian population. *Int J Obes*, in press.
- Gifford AN and Ashby CR (1996) Electrically evoked acetylcholine release from hippocampal slices is inhibited by the cannabinoid receptor agonist, WIN 55212–2, and is potentiated by the cannabinoid antagonist, SR 141716A. *J Pharmacol Exp Ther* **277**:1431–1436.
- Giuffrida A, Parsons LH, Kerr TM, de Fonseca FR, Navarro M, and Pomelli D (1999) Dopamine activation of endogenous cannabinoid signaling in dorsal striatum. *Nat Neurosci* **2**:358–363.
- Grimaldi C, Pisanti C, Laezza C, Malfitano AM, Santoro A, Vitale M, Caruso M, Notarnicola M, Iacuzzo I, Portella G, et al. (2006) Anandamide inhibits adhesion and migration of breast cancer cells. *Exp Cell Res* **312**:363–373.
- Harvie M, Howell A, Vierkant RA, Kumar N, Cerhan JR, Kelemen LE, Folsom AR, and Sellers TA (2005) Association of Gain and Loss of Weight before and after Menopause with Risk of Postmenopausal Breast Cancer in the Iowa Women's Health Study. *Cancer Epidemiol Biomarkers Prev* **14**:656–661.
- Hilairt S, Bouaboula M, Carrière D, Le Fur G, and Casellas P (2003) Hypersensitization of the orexin 1 receptor by the CB1 receptor. *J Biol Chem* **278**:23731–23737.
- Herkenham M, Lynn AB, de Costa BR, and Richfield EK (1991a) Neuronal localization of cannabinoid receptors in basal ganglia of the rat. *Brain Res* **547**:267–274.
- Herkenham M, Lynn AB, Johnson MR, Melvin LS, de Costa BR, and Rice KC (1991b) Characterization and localization of cannabinoid receptors in rat brain: a quantitative in vitro autoradiographic study. *J Neurosci* **11**:563–583.
- Herkenham M, Lynn AB, Little MD, Johnson MR, Melvin LS, de Costa BR, and Rice KC (1990) Cannabinoid receptor localization in brain. *Proc Natl Acad Sci USA* **87**:1932–1936.
- Hou Y, Yang J, Zhao G, and Yuan Y (2004) Ferulic acid inhibits endothelial cell proliferation through NO down-regulating ERK1/2 pathway. *J Cell Biochem* **93**:1203–1209.
- Hube F and Hauner H (1999) The role of TNF $\alpha$  in human adipose tissue: prevention of weight gain at the expense of insulin resistance? *Hormone Metab Res* **31**:626–631.
- Jacobsson SO, Wallin T, and Fowler CJ (2001) Inhibition of rat C6 glioma cell proliferation by endogenous and synthetic cannabinoids. Relative involvement of cannabinoid and vanilloid receptors. *J Pharmacol Exp Ther* **299**:951–959.
- Jansen EM, Haycock DA, Ward SJ, and Seybold VS (1992) Distribution of cannabinoid receptors in rat brain determined with aminoalkylindoles. *Brain Res* **575**:93–102.
- Klein TW, Newton C, Larsen K, Lu L, Perkins I, Nong L, and Friedman H (2003) The cannabinoid system and immune modulation. *J Leukoc Biol* **74**:486–496.
- Landsman RS, Burke TH, Consroe P, Roeske WR, and Yamamura HI (1997) SR141716A is an inverse agonist at the human cannabinoid CB1 receptor. *Eur J Pharmacol* **334**:R1–R2.
- Lange JHM and Kruse CG (2005) Medicinal chemistry strategies to CB1 cannabinoid receptor antagonists. *Drug Discov Today* **10**:693–702.
- Lastres-Becker I, Bizat N, Boyer F, Hantraye P, Brouillet E, and Fernandez-Ruiz J (2003) Effects of cannabinoids in the rat model of Huntington's disease generated by an intrastratial injection of malonate. *Neuroreport* **14**:813–816.
- Liu WM, Duan EK, and Cao YJ (2002) Effect of anandamide on embryo implantation in the mouse. *Life Sci* **71**:1623–1632.
- Maccarrone M, Lorenzon T, Bari M, Melino G, Finazzi-Agrò A (2000a) Anandamide induces apoptosis in human cells via vanilloid receptors. Evidence for a protective role of cannabinoid receptors. *J Biol Chem* **275**:31938–31945.
- Maccarrone M, Valensise H, Bari M, Lazzarin N, Romanini C, Finazzi-Agrò A (2000b) Relation between decreased anandamide hydrolase concentrations in human lymphocytes and miscarriage. *Lancet* **355**:1326–1329.
- Maccarrone M, Finazzi-Agrò A (2004) Anandamide hydrolase: a guardian angel of human reproduction? *Trends Pharmacol Sci* **25**:353–357.
- Maldonado R, Valverde O, and Berrendero F (2006) Involvement of the endocannabinoid system in drug addiction. *Trends Neurosci* **29**:225–232.
- Mallat A and Lotersztajn S (2006) Endocannabinoids as novel mediators of liver diseases. *J Endocrinol Invest* **29**:58–65.
- Mallet PE and Beninger RJ (1998) The cannabinoid CB1 receptor antagonist SR141716A attenuates the memory impairment produced by delta9-tetrahydrocannabinol or anandamide. *Psychopharmacology* **140**:11–19.
- Manabe Y, Toda S, Miyazaki K, and Sugihara H (2003) Mature adipocytes, but not preadipocytes, promote the growth of breast carcinoma cells in collagen gel matrix culture through cancer-stromal cell interactions. *J Pathol* **201**:221–228.
- Mazzola C, Micale V, and Drago F (2003) Amnesia induced by beta-amyloid fragments is counteracted by cannabinoid CB1 receptor blockade. *Eur J Pharmacol* **477**:219–225.
- Mechoulam R, Friede E, Di marzo V (1998) Endocannabinoids. *Eur J Pharmacol* **359**:1–18.
- Melis MR, Succu S, Mascia MS, Sanna F, Melis T, Castelli MP, and Argiolas A (2006) The cannabinoid receptor antagonist SR-141716A induces penile erection in male rats: involvement of paraventricular glutamic acid and nitric oxide. *Neuropharmacology* **50**:219–228.
- Mishima K, Nobuaki E, Hirokawa N, Fujii M, Matsumoto Y, Iwasaki K, and Fujiwara M (2001) Characteristics of learning and memory impairment induced by  $\Delta^9$  tetrahydrocannabinol in rats. *Jpn J Pharmacol* **87**:297–308.
- Naderi N, Shafagh B, Khodayar MJ, and Zarindast MR (2005) Interaction between gamma-aminobutyric acid GABA<sub>B</sub> and cannabinoid CB<sub>1</sub> receptors in spinal pain pathways in rat. *Eur J Pharmacol* **514**:159–164.
- Navarro M, Carrera MR, Fratta W, Valverde O, Cossu G, Fattore L, Chowen JA, Gomez R, del Arco I, Villanua MA, et al. (2001) Interaction between opioid and cannabinoid receptors in drug self-administration. *J Neurosci* **15**:5344–5350.
- Nesto RW (2005) Managing cardiovascular risk in patients with metabolic syndrome. *Clin Cornerstone* **7**:46–51.
- Osei-Hyiaman D, DePetrillo M, Pacher P, Liu J, Radaeva S, Bátkai S, Harvey-White J, Mackie K, Offertaler L, Wang L, et al. (2005) Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. *J Clin Invest* **115**:1298–1305.



- Perio A, Rinaldi-Carmona M, Maruani J, Barth F, Le Fur G, and Soubrie P (1996) Central mediation of the cannabinoid cue: activity of a selective CB1 antagonist, SR141716A. *Behav Pharmacol* **7**:65–71.
- Pertwee RG (2005) Inverse agonism and neutral antagonism at cannabinoid CB1 receptors. *Life Sci* **76**:1307–1324.
- Pertwee RG (2006) The pharmacology of cannabinoid receptors and their ligands: an overview. *Int J Obes (London)* **1**:S13–S18.
- Pertwee RG, Fernando SR, Nash JE, and Coutts AA (1996) Further evidence for the presence of cannabinoid CB1 receptors in guinea-pig small intestine. *Br J Pharmacol* **118**:2199–2205.
- Petitot F, Marin L, and Doble A (1996) Biochemical and pharmacological characterization of cannabinoid binding sites using [<sup>3</sup>H]SR141716A. *Neuroreport* **7**:789–792.
- Pi-Sunyer XF, Aronne LJ, Heshmati HM, Devin J, Rosenstock J, RIO-North America Study Group (2006) Effect of Rimonabant, a cannabinoid-1 receptor blocker, on weight and cardiometabolic risk factors in overweight or obese patients: a randomized controlled trial. *J Am Med Assoc* **295**:761–775.
- Portella G, Laezza C, Laccetti P, De Petrocellis L, Di Marzo V, and Bifulco M (2003) Inhibitory effects of cannabinoid CB1 receptor stimulation on tumor growth and metastatic spreading: actions on signals involved in angiogenesis and metastasis. *FASEB J* **17**:1771–1773.
- Powles T, Poole R, Shamash J, Chaplin T, Propper D, Joel S, Oliver T, and Liu WM (2005) Cannabis-induced cytotoxicity in leukemic cell lines: the role of the cannabinoid receptors and the MAPK pathway. *Blood* **105**:1214–1221.
- Ravinet Trillou C, Arnone M, Delgore C, Gonalons N, Keane P, Maffrand JP, and Soubrie P (2003) Anti-obesity effect of SR141716, a CB1 receptor antagonist, in diet-induced obese mice. *Am J Physiol* **284**:R345–R353.
- Rinaldi-Carmona M, Barth F, Healume M, Shire D, Calandra B, Congy C, Martinez S, Maruani J, Neliat G, Caput D, Ferrara P, et al. (1994) SR141716A, a potent and selective antagonist of the brain cannabinoid receptor. *FEBS Lett* **350**:240–244.
- Robson P (2005) Human studies of cannabinoids and medicinal cannabis. *Handb Exp Pharmacol* **168**:719–756.
- Rossato M, Pota FI, Ferigo M, Clari G, and Foresta C (2005) Human sperm express cannabinoid receptor Cb<sub>1</sub>, the activation of which inhibits motility, acrosome reaction, and mitochondrial function. *J Clin Endocrinol Metab* **90**:984–991.
- Sanchez C, de Ceballos ML, del Pulgar TG, Rueda D, Corbacho C, Velasco G, Galve-Roperh I, Huffman JW, Ramon y Cajal S, and Guzman M (2001) Inhibition of glioma growth in vivo by selective activation of the CB(2) cannabinoid receptor. *Cancer Res* **61**:5784–5789.
- Sarfaraz S, Afaq F, Adhami VM, and Mukhtar H (2005) Cannabinoid receptor as a novel target for the treatment of prostate cancer. *Cancer Res* **65**:1635–1641.
- Sarnataro D, Pisanti S, Santoro A, Gazzerò P, Malfitano AM, Laezza C, and Bifulco M (2006) The cannabinoid CB1 receptor antagonist rimonabant (SR141716) inhibits human breast cancer cell proliferation through a lipid raft-mediated mechanism. *Mol Pharmacol* **70**:1298–1306.
- Scheen AJ, Finer N, Hollander P, Jensen MD, Van Gaal LF, RIO-Diabetes Study Group (2006) Efficacy and tolerability of rimonabant in overweight or obese patients with type 2 diabetes: a randomized controlled study. *Lancet* **368**:1660–1672.
- Schuel H, Burkman LJ, Lippes J, Crickard K, Forester E, Pomelli D, and Giuffrida A (2002) N-Acylethanamines in human reproductive fluids. *Chem Phys Lipids* **121**:211–227.
- Segovia G, Mora F, Crossman AR, and Brochie JM (2003) Effects of CB1 cannabinoid receptor modulating compounds on the hyperkinesia induced by high-dose Levodopa in the reserpine-treated rat model of Parkinson's disease. *Mov Disord* **18**:138–149.
- Shire D, Calandra B, Delpech M, Dumont X, Kagha M, Le Fur G, Caput D, Ferrara P. (1996) Structural features of the central cannabinoid CB<sub>1</sub> receptor involved in the binding of the specific CB1 antagonist SR 141716A. *J Biol Chem* **271**:6941–6946.
- Solbrig MV, Adrian R, Baratta J, Piomelli D, and Giuffrida A (2005) A role for endocannabinoids in viral-induced dyskinetic and convulsive phenomena. *Exp Neurol* **194**:355–362.
- Spanagel R, Weiss F (1999) The dopamine hypothesis of reward: past and current status. *Trends Neurosci* **22**:521–527.
- Spano MS, Fattore L, Cossu G, Deiana S, Fadda P, and Fratta W (2004) CB1 receptor agonist and heroin, but not cocaine, reinstate cannabinoid-seeking behaviour in the rat. *Br J Pharmacol* **143**:343–350.
- Stepulak A, Siffringer M, Rzeski W, Endesfelder S, Gratopp A, Pohl EE, Bittigau P, Felderhoff-Mueser U, Kaindl AM, Bührer C, et al. (2005) NMDA antagonist inhibits the extracellular signal-regulated kinase pathway and suppresses cancer growth. *Proc Natl Acad Sci USA* **102**:15605–15610.
- Succu S, Mascia MS, Sanna F, Melis T, Argiolas A, and Melis MR (2006) The cannabinoid CB1 receptor antagonist SR141716 induces penile erection by increasing extra-cellular glutamic acid in the paraventricular nucleus of male rats. *Behav Brain Res* **169**:274–281.
- Szabo B, Dörner L, Pfreundtner C, Nörenberg W, and Starke K (1998) Inhibition of GABAergic inhibitory postsynaptic currents by cannabinoids in rat corpus striatum. *Neuroscience* **85**:395–403.
- Takahashi RN, Pampona FA, and Fernandes MS (2005) The cannabinoid antagonist SR141716A facilitates memory acquisition and consolidation in the mouse elevated T-maze. *Neurosci Lett* **380**:270–275.
- Tanda G, Pontieri FE, Di Chiara G (1997) Cannabinoid and heroin activation of mesolimbic dopamine transmission by a common mu1 opioid receptor mechanism. *Science (Wash DC)* **276**:2048–2050.
- Terranova JP, Storme JJ, Lafon N, Perio A, Rinaldi-Carmona M, Le Fur G, and Soubrie P (1996) Improvement of memory in rodents by the selective CB1 cannabinoid receptor antagonist, SR 141716. *Psychopharmacology* **126**:165–172.
- Thomas BF, Wei X, and Martin BR (1992) Characterization and autoradiographic localization of the cannabinoid binding site in rat brain using [<sup>3</sup>H]11-OH delta 9-THC-DMH. *J Pharmacol Exp Ther* **263**:1383–1390.
- Teixeira-Clerc F, Julien B, Grenard P, Tran Van Nhieu J, Deveaux V, Li L, Serriere-Lanneau V, Ledent C, Mallat A, and Lotersztajn S (2006) CB1 cannabinoid receptor antagonism: a new strategy for the treatment of liver fibrosis. *Nat Med* **12**:671–676.
- Valverde O, Karsak M, and Zimmer A (2005) Analysis of the endocannabinoid system by using CB1 cannabinoid receptor knockout mice. *Handb Exp Pharmacol* **168**:117–145.
- Van Gaal LF, Rissanen AM, Scheen A, Ziegler O, Rössner S, RIO-Europe Study Group (2005) Effects of the cannabinoid-1 receptor blocker rimonabant on weight reduction and cardiovascular risk factors in overweight patients: 1-year experience from the RIO-Europe study. *Lancet* **365**:1389–1397.
- Varga K, Wagner JA, Bridgen DT, and Kunos G (1998) Platelet- and macrophage-derived endogenous cannabinoids are involved in endotoxin-induced hypotension. *FASEB J* **12**:1035–1044.
- Walker JM, Hohmann AG (2005) Cannabinoid mechanisms of pain suppression. *Handb Exp Pharmacol* **168**:509–554.
- Wang L, Liu J, Harvey-White J, Zimmer A, and Kunos G (2003) Endocannabinoid signaling via cannabinoid receptor 1 is involved in ethanol preference and its age-dependent decline in mice. *Proc Natl Acad Sci USA* **100**:1393–1398.
- Wilson RI, Nicoll RA (2002) Endocannabinoids signaling in the brain. *Science (Wash DC)* **296**:678–682.
- Yang WS, Lee WJ, Funahashi T, Tanaka S, Matsuzawa Y, Chao CL, Chen CL, Tai TY, and Chuang LM (2001) Weight reduction increases plasma levels of an adipose-derived anti-inflammatory protein, adiponectin. *J Clin Endocrinol Metab* **86**:3815–3819.
- Yoon S, and Seger R (2006) The extracellular signal-regulated kinase: multiple substrates regulate diverse cellular functions. *Growth Factors* **24**:21–44.

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